

Isolation and prevalence of *Campylobacter* species in cattle from Sokoto State, Nigeria

Mohammed D. Salihu⁽¹⁾, Junaidu U. Abdulkadir⁽¹⁾, Steve I. Oboegbulem⁽¹⁾,
Godwin O. Egwu⁽²⁾, Abdullahi A. Magaji⁽¹⁾, Mohammed Lawal⁽¹⁾
& Yahaya Hassan⁽¹⁾

Summary

The prevalence of *Campylobacter* spp. in cattle in Sokoto State, Nigeria, was determined. The number of samples collected totalled 976, of which 126 (12.9%) yielded *Campylobacter* spp. The species of *Campylobacter* isolates from this study were as follows: *C. jejuni* (65.1%), *C. coli* (23.0%), *C. lari* (7.9%), *C. hyointestinalis* (3.2%) and *C. fetus* (0.8%). A total of 172 strains of *Campylobacter* spp. were identified from the positive samples due to identification of more than a single strain (spp.) from a single sample. The strains identified were *C. jejuni* (62.8%), *C. coli* (25.0%), *C. lari* (8.1%), *C. hyointestinalis* (2.9%) and *C. fetus* (1.2%). More than one species of *Campylobacter* was identified in 36.5% of the positive samples. The biotyping in this study revealed *C. jejuni* biotype I (34.3%) as the most common *C. jejuni* biotype, while *C. jejuni* biotype IV (15.7%) was the *C. jejuni* biotype that was least frequently isolated. However, the most frequently isolated *C. coli* biotype was biotype I (72.1%) and all the isolates of *C. lari* were biotype I.

Keywords

Biotype, *Campylobacter*, Cattle, Isolation, Nigeria, Prevalence, Public health, Sokoto.

Isolamento e prevalenza di *Campylobacter* in allevamenti bovini nello stato di Sokoto in Nigeria

Riassunto

Nel presente lavoro viene esaminata la prevalenza di *Campylobacter* spp. in allevamenti bovini nello stato di Sokoto in Nigeria. Su 976 campioni, 126 (12,9%) sono risultati positivi per *Campylobacter* spp. Sono stati isolati: *C. jejuni* (65,1%), *C. coli* (23,0%), *C. lari* (7,9%), *C. hyointestinalis* (3,2%) e *C. fetus* (0,8%). Sono stati identificati 172 ceppi di *Campylobacter* spp. da campioni positivi e più ceppi (spp.) da un singolo campione. I ceppi identificati sono stati *C. jejuni* (62,8%), *C. coli* (25,0%), *C. lari* (8,1%), *C. hyointestinalis* (2,9%) e *C. fetus* (1,2%). Nel 35,6% di campioni positivi è stata identificata più di una specie di *Campylobacter*. La biotipizzazione ha evidenziato tra i più diffusi *C. jejuni* biotipo I (34,3%) e *C. jejuni* biotipo IV (15,7%), il *C. jejuni* biotipo è risultato quello isolato con meno frequenza. Il biotipo di *C. coli* isolato con più frequenza è stato il biotipo I (72,1%). Tutti gli isolamenti di *C. lari* sono risultati del biotipo I.

Parole chiave

Biotipo, Bovino, *Campylobacter*, Isolamento, Nigeria, Prevalenza, Salute pubblica, Sokoto.

(1) Department of Veterinary Public Health and Animal Production, Faculty of Veterinary Medicine, Usmanu Danfodiyo University, P.M.B. 2254, Sokoto, Sokoto State, Nigeria
mdsal70@yahoo.com

(2) Department of Veterinary Microbiology and Pathology, Faculty of Veterinary Medicine, Usmanu Danfodiyo University, P.M.B. 2254, Sokoto, Sokoto State, Nigeria

Introduction

Campylobacter species are common bacterial pathogens that cause gastroenteritis in humans, both in industrialised and developing countries (9, 32, 38). Members of the genus *Campylobacter* have long been recognised as a cause of septic abortion in both cattle and sheep (4), but the development and improvement of *Campylobacter* selective culture media lead to the recognition that *Campylobacter* can be an aetiological agent of human gastroenteritis (34).

A variety of *Campylobacter* subspecies have been isolated from healthy and diseased cattle (3). *Campylobacter* have been reported to cause enteritis in calves (1, 2). *Campylobacter jejuni* ssp. *jejuni* and *C. hyointestinalis* have occasionally been reported to cause bovine abortion (12, 36) and *C. jejuni* ssp. *jejuni* has been isolated in bovine mastitis (29). *Campylobacter* species have been isolated from the faeces of healthy cattle (3, 5, 18, 19, 20, 24). Although the chicken is the species most frequently identified as a reservoir of bacteria responsible for human infection, major outbreaks have been recorded from contaminated or inadequately pasteurised milk (16, 28, 31). Studies have reported an association between *Campylobacter* infection in humans and contact with cattle (3, 18, 23, 25, 30, 35).

Direct-contact exposure to bovine faeces and ingestion of unpasteurised bovine milk are well documented causes of outbreaks of campylobacteriosis (15, 33). A high degree of genetic relatedness between the *Campylobacter* from cattle and humans in the same geographical area has been reported by Fitzgerald *et al.* (17). Given the potential linkage between *Campylobacter* spp. harboured by cattle and human disease, the present study was conducted to determine the prevalence and distribution of a variety of *Campylobacter* spp. in cattle in Sokoto State.

Materials and methods

The study was conducted between November 2007 and January 2009. The prevalence of the

infection was estimated on the basis of expected prevalence of 50%, with 5% as the maximum acceptable error and 95% the confidence interval. The choice of 50% expected prevalence was due to a lack of knowledge of the true prevalence of *Campylobacter* infection in cattle in the state (Sokoto) and the country (Nigeria). A total of 976 rectal swabs were collected from cattle across the state and transported to the laboratory within 6 h of the time of sampling. At the laboratory, each individual swab was inoculated into modified charcoal cefoperazone deoxycholate agar (mCCDA) medium (Oxoid, CM739) supplemented with cefoperazone and amphotericin B (Oxoid, SR155) for selective isolation of *Campylobacter* spp. at 42°C for 48 h to 96 h in an anaerobic jar containing a microaerophilic generating sachet (Campygen) (Oxoid, CN35A).

From each plate, up to five colonies with colonial morphology consistent with thermophilic *Campylobacter* spp. were streaked for isolation on blood agar and incubated for 24 h at 42°C. Presumptive thermophilic *Campylobacter* colonies were then suspended in proteose peptone glycerol (10%) and stored at -70°C for subsequent species identification and biotyping.

Campylobacter isolates were cultured on Columbia agar plates containing 5% sheep blood in a microaerophilic condition (Campygen, Oxoid, CN35A). All isolates were characterised using the standard *Campylobacter* phenotypic identification procedure described by Atabay and Corry (3). Biotyping of the isolates was performed using the extended biotyping scheme described by Lior (27).

Results

The major phenotypic characteristics of the isolates in this study were typical of thermophilic *Campylobacter* spp. All the isolates demonstrated spiral or curved rod morphology in Gram staining, the isolates revealed oxidase and catalase activity and reduced nitrate. Hippurate was hydrolysed by suspected *C. jejuni* isolates.

A total of 126 samples (12.9%) were positive for *Campylobacter* isolations. *C. jejuni* was isolated in 82 (65.1%) of the positive samples, *C. coli* in 29 samples (23.0%) and *C. lari* in 10 (7.9%) of the positive samples. *C. hyointestinalis* and *C. fetus* were isolated in 4 (3.2%) and 1 (0.8%) of the positive samples, respectively, as shown in Table I. Overall, 172 strains of the *Campylobacter* spp. were identified due to isolation of more than a single strain (species) from a single sample. Approximately 36.5% of the positive samples yielded more than a single species of *Campylobacter*. The strains identified were as follows (Table I):

- *C. jejuni* 108 (62.8%)
- *C. coli* 43 (25.0%)
- *C. lari* 14 (8.1%)
- *C. hyointestinalis* 5 (2.9%)
- *C. fetus* 2 (1.2%).

Table I
Percentage isolation of *Campylobacter* spp.

Species	No. positive (%)	No. of strains isolated (%)
<i>Campylobacter jejuni</i>	82 (65.1%)	108 (62.8%)
<i>Campylobacter coli</i>	29 (23.0%)	43 (25.0%)
<i>Campylobacter lari</i>	10 (7.9%)	14 (8.1%)
<i>Campylobacter hyointestinalis</i>	4 (3.2%)	5 (2.9%)
<i>Campylobacter fetus</i>	1 (0.8%)	2 (1.2%)

The isolates were biotyped as described by Lior (27); the biotyping revealed that *C. jejuni* biotype I 37 (34.3%) was the most common *C. jejuni* biotype in this study, while *C. jejuni* biotype IV 17 (15.7%) was the least frequently *C. jejuni* biotype isolated. However, the most frequently isolated *C. coli* biotype was biotype I 31 (72.1%); all isolates of *C. lari* were biotype I (Table II).

Discussion

The occurrence of human *Campylobacter* gastroenteritis has largely been attributed to the consumption of contaminated food animal products (4), especially poultry, because of the high prevalence of *Campylobacter* in these animals (10, 11, 22). There is evidence that

suggests other vehicles such as red meat, environmental water and unpasteurised milk as important sources of these organisms (21, 33).

Table II
Biotypes of the thermophilic *Campylobacter* isolates

<i>Campylobacter</i>	Biotypes	Isolation rate (%)
<i>Campylobacter jejuni</i>	I	37 (34.3%)
	II	26 (24.1%)
	III	28 (25.9%)
	IV	17 (15.7%)
<i>Campylobacter coli</i>	I	31 (72.1%)
	II	12 (27.9%)
<i>Campylobacter lari</i>	I	14 (100%)
	II	0 (00.0%)

The isolation conditions in this study were developed for the isolation of thermophilic campylobacters (*C. jejuni*, *C. coli* and *C. lari*). This may bias detection in favour of thermophilic campylobacters (3, 4, 6, 14). The prevalence of thermophilic campylobacters in this study was 12.9% and is in agreement with previous studies in which prevalence of campylobacters in cattle ranged from 0.8% to 46.7%, depending on the method of isolation, season, age of animal and sample size (4, 8, 19, 23, 32, 37).

The concurrent excretion frequency in this study (36.5%) is higher than the 9% rate reported by Bae *et al.* (4) and 24% reported by Inglis *et al.* (25). Considering the composition of isolation media and the isolation conditions in this study, the isolation of *C. hyointestinalis* is however possible but was isolated at a lower rate in this study. This is in agreement with the observations of Atabay and Corry (3), Busato *et al.* (7), Giacoboni *et al.* (19) and Inglis *et al.* (25). The isolation of thermotolerant *C. fetus* in this study was fortuitous; some investigators have reported the isolation of similar atypical *C. fetus* strains from raw milk and humans (13, 26, 39).

The most common biotype of *C. jejuni* in this study was biotype I which accounts for 34.3% of the total *C. jejuni* isolates. This observation

was in agreement with that of Baserisalehi *et al.* (5) who observed more of the *C. jejuni* biotype I than any other *C. jejuni* biotypes. The common *C. coli* biotype in this study was biotype I. The identification of these biotypes from cattle in the state is of serious public

health importance, since these biotypes have been implicated as the cause of disease in humans.

The results of this study demonstrate that *C. jejuni* is widely distributed among cattle in Sokoto State.

References

1. Al-Mashat R.R. & Taylor D.J. 1980. Production of diarrhoea and dysentery in experimental calves by feeding pure cultures of *Campylobacter fetus* subsp. *jejuni*. *Vet Rec*, **107**, 459-464.
2. Al-Mashat R.R. & Taylor D.J. 1980. Production of enteritis in calves by the oral inoculation of pure cultures of *Campylobacter fetus* subsp. *intestinalis*. *Vet Rec*, **112**, 54-58.
3. Atabay H.L. & Corry J.E.L. 1998. The isolation and prevalence of campylobacters from the dairy using a variety of methods. *J Appl Microbiol*, **84**, 733-740.
4. Bae W., Kaya K.N., Hancock D.D., Call D.R., Park Y.H & Besser T.E. 2005. Prevalence and antimicrobial resistance of thermophilic *Campylobacter* spp. from cattle farms in Washington State. *J Appl. Environ Microbiol*, **71** (1), 169-174.
5. Baserisalehi M., Bahadur N. & Kapadnis B.P. 2007. Isolation and characterization of *Campylobacter* spp. from domestic animals and poultry in south of Iran. *Pakistan J Biol Sci*, **10** (9), 1519-1524.
6. Baylis C.L., MacPhee S., Martin, K.W., Humphher T.J. & Betts R.P. 2000. Comparison of three enrichment media for the isolation of *Campylobacter* spp. from foods. *J Appl Microbiol*. **89**, 884-891.
7. Busato A., Hofer D., Lentze T., Gaillard C. & Burnens A. 1999. Prevalence and infection risks of zoonotic enteropathogenic bacteria in Swiss cow-calf farms. *Vet Microbiol*, **69**, 251-263.
8. Cabrita J., Rodrigues J., Braganca F., Morgado C., Pires I. & Goncalves A.P. 1992. Prevalence, biotypes, plasmid profile and antimicrobial resistance of *Campylobacter* isolated from wild and domestic animals from northeast Portugal. *J Appl Bacteriol*, **73**, 279-285.
9. Coker A.O., Isokpehi R.D., Thomas B.N., Amisu K.O. & Obi C.L. 2002. Human campylobacteriosis in developing countries. *Emerg Infect Dis*, **8**, 237-244.
10. Corry J.E.L. & Atabay H.I. 2001. Poultry as a source of *Campylobacter* and related organisms. *J Appl Microbiol*, **90**, 96S-114S.
11. Deming M.S., Tauxe R.V., Blake P.A., Dixon S.E., Fowler B.S., Jones T.S., Lockamy E.A., Patton C.M. & Sikes R.O. 1987. *Campylobacter* enteritis at a university: transmission from eating chickens and from cats. *Am J Epidemiol*, **126**, 526-534.
12. Diker K.S., Diker S. & Ozlem M.B. 1990. Bovine diarrhea associated with *Campylobacter hyointestinalis*. *Zentralbl Veterinarmed B*, **37**, 158-160.
13. Edmond P., Patton C.M., Barrett T.J., Morris G.K., Steigerwalt A.G. & Brenner D.J. 1985. Biochemical and genetic characteristics of atypical *Campylobacter fetus* subsp. *fetus* strain isolated from humans in the United States. *J Clin Microbiol*, **21**, 936-940.
14. Engberg J., On S.L.W., Harrington C.S. & Gerner-Smidt P. 2000. Prevalence of *Campylobacter*, *Arcobacter*, *Helicobacter*, and *Sutterella* spp. in human fecal samples as established by a reevaluation of isolation methods for campylobacters. *J Clin Microbiol*, **38**, 286-291.
15. Evans M.R., Roberts R.J., Ribeiro C.D., Gardner D. & Kembrey D. 1996. Milk-borne *Campylobacter* outbreak following an educational farm visit. *Epidemiol Infect*, **117**, 457-462.
16. Fahey T., Morgan D., Gunneburg C., Adak G.K., Majid F. & Kaczmarek E. 1995. An outbreak of *Campylobacter jejuni* enteritis associated with failed milk pasteurisation. *J Infect*, **31**, 137-143.
17. Fitzgerald C., Stanley K., Andrew S & Jones K. 2001. Use of pulsed-field gel electrophoresis and flagellin gene typing in identifying clonal groups of *Campylobacter jejuni* and *Campylobacter coli* in farms and clinical environments. *Appl Environ Microbiol*, **67**, 1429-1436.
18. Garcia M.M., Lior H., Stewart G.M., Ruckerbauer J.R., Trudel R. & Skljarevski A. 1985. Isolation, characterization, and serotyping of *Campylobacter jejuni* and *Campylobacter coli* from slaughter cattle. *Appl Environ Microbiol*, **49**, 667-672.
19. Giacoboni G.L., Itoh K., Hirayama K., Takahashi E. & Mitsuoka T. 1993. Comparison of fecal *Campylobacter* in calves and cattle of different ages and areas in Japan. *Jap J Vet Med Sci*, **55**, 555-559

20. Grau F.H. 1988. *Campylobacter jejuni* and *Campylobacter hyointestinalis* in the intestinal tract on the carcasses of calves and cattle. *J Food Protect*, **51**, 857-861.
21. Hanninen M.L., Pajarre S., Klossner M.L. & Rautelin H. 1998. Typing of human *Campylobacter jejuni* isolates in Finland by pulsed-field gel electrophoresis. *J Clin Microbiol*, **36**, 1787-1789.
22. Harris N.V., Thompson D., Martin C. & Nolan C.M. 1986. A survey of *Campylobacter* and other bacterial contaminants of pre-market chicken and retail poultry and meats, King County, Washington. *Am J Publ Health*, **76**, 401-406.
23. Hoar B.R., Atwill E.R., Elmi C. & Farver T.B. 2001. An examination of risk factors associated with beef cattle shedding pathogens of potential zoonotic concern. *Epidemiol Infect*, **127**, 147-155.
24. Humphrey T.J. & Beckett P. 1987. *Campylobacter jejuni* in dairy cows and raw milk. *Epidemiol Infect*, **98** (3), 63-69.
25. Inglis G.D., Kalischuk L.D. & Busz H.W. 2003. A survey of *Campylobacter* species shed in faeces of beef cattle using polymerase chain reaction. *Can J Microbiol*, **49**, 655-661.
26. Klein B.S., Vergeront J.M., Blaser M.J., Edmonds P., Brenner D.J., Janssen D. & Davis J.P. 1986. *Campylobacter* infection associated with raw milk: an outbreak of gastroenteritis due to *Campylobacter jejuni* and thermotolerant *Campylobacter fetus* subsp. *fetus*. *J Am. Med. Assoc*, **255**, 361-364.
27. Lior H. 1984. New, extended biotyping scheme for *Campylobacter jejuni*, *Campylobacter coli* and '*Campylobacter lariidis*'. *J Clin Microbiol*, **20**, 636-640.
28. Morgan D., Gunneberg C., Gunnell D., Healing T.D., Lamerton S., Soltanpoor N., Lewis D.A. & White D.G. 1994. An outbreak of *Campylobacter* infection associated with the consumption of unpasteurised milk at a large festival in England. *Eur J Epidemiol*, **10**, 581-585.
29. Morgan G., Chadwick P., Lander K.P. & Gill K.P. 1985. *Campylobacter jejuni* mastitis in a cow: a zoonoses-related incident. *Vet Rec*, **116**, 111.
30. Nielsen E.M. 2002. Occurrence and strain diversity of thermophilic campylobacters in cattle of different age groups in dairy herds. *Letters Appl Microbiol*, **35**, 85-89.
31. Pearson A.D. & Healing T.D. 1992. The surveillance and control of *Campylobacter* infection. *Commun Dis Rep CDR Rev*, **2**, R133-R139.
32. Pebody R.G., Ryan M.J. & Wall P.G. 1997. Outbreaks of *Campylobacter* infection: rare events for a common pathogen. *Commun Dis Rep CDR Rev*, **7**, R33-R37.
33. Peterson M.C. 2003. *Campylobacter jejuni* enteritis associated with consumption of raw milk. *J Environ Health*, **65**, 20-21.
34. Skirrow M.B. 1977. *Campylobacter* enteritis: a 'new' disease. *Br Med J*, **2**, 9-11.
35. Stanley K. & Jones K., 2003. Cattle and sheep farms as reservoirs of *Campylobacter*. *J Appl Microbiol*, **94**, 1045-1135.
36. Van Donkersgoed J., Janzen E.D., Chirino-Trejo M., Berry C., Clark E.G. & Haines D.M. 1990. *Campylobacter jejuni* abortions in two beef cattle herds in Saskatchewan. *Can Vet J*, **31**, 373-377.
37. Wesley I.V., Wells S.J., Harmon K.M., Green A., Schroeder-Tucher L., Glover M. & Siddique I. 2000. Fecal shedding of *Campylobacter* and *Arcobacter* spp. in dairy cattle. *Appl Environ Microbiol*, **66**, 1994-2000.
38. World Health Organization (WHO) 2000. *Campylobacter*. Fact sheet No. 255, November 2000. WHO, Geneva, 1 p (www.who.int/mediacentre/factsheets/fs255/en/ accessed on 1 August 2009).
39. Woo P.C., Leung K.W., Tsoi H.W., Wong S.S., Teng J.L. & Yuen K.Y. 2002. Thermotolerant *Campylobacter fetus* bacteraemia identified by 16S ribosomal RNA gene sequencing: an emerging pathogen in immunocompromised patients. *J Med Microbiol*. **51**, 740-746.